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An Approach to Ultrasonic Risk Assessment and an  
Examination of Selected Experimental Studies

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## INTRODUCTION

It has been more than a decade since the first major efforts to assess the risk of ultrasonic energy have been made (1, 2). Since then there have been numerous reviews and assessments (3-10). And, even today, there are projects which have as their aim an assessment of the health risks from diagnostic ultrasound (11-13). And yet, the basic status regarding risk assessment has remained unchanged during this time viz., the studies necessary to support a reliable assessment of the risks associated with human exposure to ultrasound have not been done. So, rather than present another broad discussion of ultrasonic risk assessment, it seemed to be much more reasonable to focus upon one ultrasonically induced biological observation, viz., fetal weight reduction. But, prior to a discussion of ultrasonically induced fetal weight reduction, let us first consider an approach to risk assessment and then a view of the general trends as they relate to diagnostic level, ultrasonic biological effects. Here, the effect of ultrasound on immunological function and on sister chromatid exchange frequency will be considered as examples.

## APPROACH TO RISK ASSESSMENT

With our current understanding of ultrasonically induced biological effects, it is difficult to argue against statements such as "diagnostic ultrasound is not harmful to the conceptus." Experimental studies can not be used to prove diagnostic ultrasound safe. Rather, what such studies

will provide, if properly planned and executed, are data which will aid in the overall assessment of risk associated with exposure to ultrasound. Safe implies the absence of an effect, not involving risk, or the like. It simply is not possible to prove that ultrasound, or for that matter any agent, does not produce any effect at the levels employed diagnostically. Also, the actual use of the word safe is vague since it almost never refers to the absence of an effect. A more useful and workable approach is to examine the "risk" associated with ultrasonic exposure.

Some thirty-five years after the Curies discovered piezoelectricity in 1880 (14), the first use of ultrasonic energy was developed wherein underwater acoustic echoes were bounced off of submerged objects (15, 16). During the course of this work, the first reported observation was made that ultrasonic energy had a lethal effect upon small aquatic animals (17). The first extensive investigation of the phenomena confirmed that ultrasonic energy could kill small fishes and frogs within a minute or two (18). In perhaps the first review paper of ultrasonically induced biological effects (19) the physical, chemical and biological effect of ultrasound were evaluated wherein effects on cells, isolated cells, bacteria and tissues were summarized with a view towards identifying the responsible mechanism. The ultrasonic exposure conditions of this early work were not well characterized but the intensity levels were undoubtedly quite high.

In the early pioneering studies where the ultrasonic

exposure conditions were carefully controlled and specified, sciatic nerve paralysis was easily produced in the frog (20, 21) and lesions were produced in central nervous system tissue (22). In addition, high intensity ultrasound was employed to produce lesions in adult cat and rat brain (23-27); adult rat and neonatal mouse spinal cord (22, 28, 29); adult frog muscle (30, 31); rabbit blood vessel (32); rabbit kidney and testicle (33); and rabbit ocular tissue (34-36). The ultrasonic intensities were very much higher than those utilized in diagnostic ultrasound. Mostly, these studies caused rather severe tissue damage, but these studies have been extremely important in the elucidation of fundamental interaction processes. In terms of risk assessment, these studies support the view that the currently employed ultrasonic exposure conditions will more than likely not produce acute, gross irreversible damage.

These high intensity studies further aid in recognizing the important fact that, at sufficient energy levels, ultrasound is capable of destroying biological material. Therefore, an approach to the question of assessing the risk from ultrasound is as follows:

- (1) what biological systems are most sensitive to ultrasound?
- (2) what exposure levels imposed a significant risk on these systems?

Unfortunately, this approach has its difficulties. How does one determine significant risk? Usually, what is meant by

significant risk is that risk which is greater than some upper limit of acceptability. One usually then employs a benefit vs risk concept which, in principal, is quite understandable but, in practice, poses severe implementation problems.

Another important consideration with respect to risk assessment is the extent to which ultrasound is used. While no statistically based survey has been conducted to document the extent to which ultrasound has been used over the past decade, a number of indicators do, however, support the view that its use is increasing and that a large fraction of the human population will eventually be exposed, especially in utero.

As listed in Table 1, in 1971, the Food and Drug Administration's Bureau of Radiological Health surveyed 301 out of 6306 short-term general hospitals in the United States and found that 12 percent of the hospitals used diagnostic ultrasound (37). The same Federal agency reported on the conduct of its 1974 hospital survey in which 35 percent of the surveyed hospitals used diagnostic ultrasound (38, 39). This represented an almost 200 percent increase in use between 1971 and 1974, or an annualized increase of 43 percent during this time. The survey (38, 39) further showed that an estimated 16 percent of the obstetric services in the United States used diagnostic ultrasound and that about one-third of all U.S. births for 1974 were delivered in these hospitals. Additionally, it was estimated that 470,000 pregnant women were exposed to diagnostic ultrasound in 1974

with about 35-40 percent of these women being examined more than once.

An international mail survey showed that between 1963 and 1971, there was an average annual increase in use of clinical ultrasound of approximately 10 percent (40). And, in the United Kingdom, it had been estimated that in the early 1970's the number of ultrasonic diagnostic examinations was doubling every three years (41), thus representing an annual 26 percent increase.

While more current use information is not available, sales growth information does indicate that in 1976, the ultrasonic industry annual dollar sales was around \$30 million (42) and for the following year about \$40 million (43). In the next four years, estimates of annual sales were \$50, \$79, \$170, and \$214 million (10). Therefore, an increase from \$30 to \$214 million from 1975 to 1980 represents an average annual increase in sales of approximately 48 percent.

To summarize, if one were to assume that at least one-half of all pregnant women are currently being examined with ultrasound and, additionally, were to assume that its use is increasing at an annual rate of between 10 and 25 percent, then within a few years, virtually every fetus could be diagnostically examined with ultrasound.

#### GENERAL TREND

Recent reviews (6, 9, 10, 44-49) of the ultrasonic bioeffect literature appear to suggest that as more sensitive

biological endpoints are studied, the ultrasonic exposure parameters required to produce measurable effects appear to decrease. This is supported by published work (50-57) which describes ultrasonically induced biological effects from ultrasonic diagnostic equipment, that is, ultrasonic energy from a commercial diagnostic device altered the biological system.

It should be noted that at the lower, sometimes diagnostic, levels of ultrasound, there is no consistent indication of the specific ultrasonically induced biological alteration. For the two examples to be discussed, immunological function and sister chromatid exchange, there are considerable conflicting experimental data as to whether these alterations are ultrasonically induced.

In the first study by Anderson and Barrett (50), which reported an effect of ultrasound on immunological function the splenic area of mice was exposed to ultrasound via a pulsed diagnostic device and it was reported that ultrasound exposure reduced the hemagglutinin and hemolysin response after an injection of sheep erythrocytes. However, another laboratory (58) could not reproduce these effects and reported that ultrasound had no significant effect on the hemoglutinin and hemolysin response in mice exposed to ultrasound under very similar conditions to that used in the earlier study (50). Further, in another experiment (58) at higher spatial average, temporal average intensities, no effect was produced.

In a second study, Anderson and Barrett (55) again

exposed mice to diagnostic levels of ultrasound, but this time to the liver area. They reported that ultrasound caused a depression of phagocytosis as measured by carbon clearance, and a depression in the ability of peritoneal macrophages from sonicated animals to phagocytose bacteria. These studies are very interesting from the standpoint that diagnostic levels of ultrasound may affect immune function. There are, however, many problems in translating these findings to the clinical situations, such as the large difference in area of sonicated vs whole body/organ size in mouse vs man.

Liebeskind and colleagues (51) reported that diagnostic levels of ultrasound increased immunoreactivity to antinucleoside antibodies in cultured cells. This type of effect usually suggests unwinding of the DNA helix, or single-strand breaks but the authors were not able to identify DNA strand damage. While this effect is sometimes referred to as an immune effect, it is not an effect on the immune system, but the use of an immune technique as an experimental tool.

Within the last five years, a number of experimental studies have been conducted to examine the effect of ultrasonic energy on sister chromatid exchange (SCE) frequency, an indicator of chromosome damage but of which the biological significance is unclear. Liebeskind's study (52) has received, perhaps, the greatest attention since the study indicated an increase in human lymphocyte SCE's from a diagnostic ultrasound device. In another study from the same



authors (51), they reported no change in SCE's in a different cell type. There has been only one other positive observation (58) of increased SEC's; here in human lymphocytes at diagnostic levels. But, there have been at least eight other studies (51, 58-64), some at diagnostic levels, some at levels much higher than therapeutic levels which have reported no increase in SCE's. Therefore, it is necessary to put into perspective all of the reported findings, both positive and negative.

One must be caution in assessing the significance of biological alterations such as the immune disturbances or increased SCE's as discussed above, or other such observations. They are simply observations, and whether or not ultrasound will be shown to represent a significant risk will depend upon the types of effects observed, the exposure levels at which these effects occur, their dose-effect responses, and the assessment of the mechanism from dose effect data. These studies are useful, however, in pointing to potentially useful hazardous or potentially therapeutic interactions of ultrasound and to help understand how ultrasound interacts with biological tissue.

#### FETAL WEIGHT STUDIES

Over the last few years, and especially within the past year or so, experimental observations have been made which suggest that subtle effects are occurring to rodent embryos and fetuses when exposed to ultrasound in utero. The balance of the article selectively examines ultrasonically induced

fetal weight reduction in experimental animals. In choosing this topic, an attempt has been made to approach the assessment of risk from ultrasound for this single biological endpoint. It is not known whether this biological system is very sensitive (in a chemical and physical sense) to ultrasonic energy but, clearly, it is sensitive in an emotional and political sense. Therefore, it behooves us to understand the experimental data which shows that ultrasonic energy does influence fetal weight when the system is exposed to ultrasound in utero.

One of the earliest studies that suggested that in utero ultrasonic irradiation affected prenatal growth and development was reported in experimental animals about eight years ago (65). Time-mated mice received continuous wave (1 MHz) ultrasound on the eighth day of gestation under well controlled and documented exposure conditions (66). The fetuses were weighed on the eighteenth day of gestation and a statistically significant weight reduction of up to 17.5 percent relative to the sham was observed. Two hundred and seventy two litters (2866 fetuses) were examined in seven separate ultrasound groups, including a sham group. The spatial average, time average intensity ranged from 0.5 to 5.5 W/cm<sup>2</sup> and the exposure time ranged from 10 to 300 seconds as listed in Table 2.

A detail account of the initial finding (65) and an extension of the data analysis (67) showed that a dose-effect response was observed. Here the dose-effect response of the exposure condition versus fetal weight was developed by

defining the dose parameter  $I^2t$ , where  $I$  is the exposure intensity ( $\text{W}/\text{cm}^2$ ) and  $t$  is the exposure time (sec) as listed in Table 2 for each of the seven exposure conditions.

There is a basis for the  $I^2t$  dose parameter in the ultrasonic literature and in the literature of other energy forms. Threshold ultrasonic dosages for structural changes in the adult mammalian central nervous system results in a mathematical dependency between the ultrasonic intensity,  $I$ , and the exposure duration,  $t$  (24, 26, 68-70) which is described by the product of  $I^2$  and  $t$  equalling a constant value. In other words, if  $I$  equals 20  $\text{W}/\text{cm}$  ( $I^2=400$ ) and  $t$  equals 0.5 sec for the threshold of an ultrasonically induced change, then  $I^2t=200$ . This same biological threshold would occur under the ultrasonic exposure conditions in which  $I$  equals 10  $\text{W}/\text{cm}$  ( $I^2=100$ ) and  $t$  equals 2.0 sec, that is  $I^2t=200$  again.

A similar type of  $I^2t$  dependency has been observed for ultrasonically induced hind limb paralysis of neonatal mice (26, 71), threshold focal lesions in cat liver (72) as well as focal lesions in the rabbit liver, kidney and teste (33). In comparison to other forms of energy, a similar dose dependency has been observed for mammary neoplasms at low ionizing radiation doses wherein two x-ray secondary particles (produced by a single neutron) are required to elicit the effect (73). In photochemical and photobiological studies, at high energy concentrations, biophotonic excitation (two photons required to produce the effect) has been observed (74).

What does an  $I^2t$  type of dependency for a biological effect mean? Basically, it means that two energy events are required to produce the biological effect. However, there is not enough fundamental information available at this time to speculate as to whether or not the reduced fetal weight observations reported herein can be explained by this dosimetric model. Nevertheless, such a model begins to provide a basis for extrapolating biological effect observations and such an extrapolation will be done shortly in terms of assessing the potential for ultrasonically induced fetal weight reduction in humans based upon the mouse data. Thus the following fetal weight observations will be presented in terms of their  $I^2t$  dependency.

The observation that in utero ultrasonic exposure can cause reduction in the mouse fetuses has been confirmed by two other research groups using two different strains of mice (75-76). For the data listed in Table 3 (75), relatively high level, pulsed ultrasound conditions were employed. Here the mice were exposed to ultrasound on the eighth day of gestation and the fetuses were individually weighed on the eighteenth day of gestation. A statistically significant 18.8 percent weight reduction was observed for spatial peak, temporal average intensities above  $50 \text{ W/cm}^2$  (spatial peak, temporal peak intensity of  $1936 \text{ W/cm}^2$  and exposure time of 20 sec) when irradiated on day eight of gestation. At and below a spatial peak, temporal average intensity of  $45 \text{ W/cm}^2$  no statistically significant change in the fetal weight was observed.

The data listed in Table 4 (76) shows mean fetal weight reductions which range up to 25 percent relative to the sham under continuous wave (2 MHz) exposure conditions at a spatial average, temporal average intensity of  $1 \text{ W/cm}^2$ , for exposure times up to 200 sec at gestation ages of zero, 7 and 12. The fetuses were individually weighed on the seventeenth day of gestation. These results compare most directly with the previous finding (65, 67) in which similar mean fetal weight reductions were observed.

Figure 1 graphically summarizes in a unified way the three published studies (67, 75, 76) which reported statistically significant effects of fetal weight reduction from in utero exposure to ultrasound. Here the data from Tables 2, 3 and 4 are represented by the percentage weight change (re: sham) as a function of the calculated dose parameter  $I^2 t$ . All three studies graphically show that as the value of  $I^2 t$  increases the fetal weight (re: sham) decreases.

#### DOSE-EFFECT STUDIES

Dose-effect studies are invaluable for assessing risk (6). Too often, a report appears in which a biological effect is reported under a single ultrasonic exposure condition. When the exposure condition arises from a diagnostic device, we tend to question whether the reported observation is real, or whether the experimental set-up produced extraordinary conditions to elicit the effect. But, in either case, there is a tendency to suggest that

diagnostic exposure conditions represent a risk to the patient. When the exposure condition is at levels much in excess of diagnostic conditions, we tend to discount their applicability to the clinical situation. The overall problem is that non dose-effect studies are quite difficult to apply to assessing the risk. They do, however, identify biological endpoint to which dose-effect experimental regimes should then be applied.

Consider the mouse dose-effect fetal weight data shown in Figure 1. If we were to apply this dose-effect curve to a clinical exposure condition for purposes of assessing risk, let us first examine the upper value of the dose parameter  $I^2t$  for static pulse echo scanners (77). For a single pulse, the spatial peak, pulse average intensity is about  $300 \text{ W/cm}^2$  and the exposure time, here the pulse duration, is about 1 microsecond, yielding an  $I^2t$  around 0.09. For the time average case, the spatial peak, temporal average intensity is about  $200 \text{ mW/cm}^2$  and the exposure time, here the length of the exam for maximum effect, is about 30 minutes, yielding an  $I^2t$  around 72. Of course, this latter case would require examining the same tissue volume for the entire length of time, which might not be the situation with static pulse echo scanner but is quite possible for a Doppler fetal monitor wherein the spatial peak intensity is about  $75 \text{ mW/cm}^2$ . For an exposure time of one hour, the  $I^2t$  dose parameter calculates to be about 20. The point to be made is that with a dose-effect model, one is in a better position of examining what might be the effect under clinical conditions. The

model would have to be validated for such applicability, of course. There is a long way to go with respect to ultrasound.

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REFERENCES

1. Reid JM, Sikov MR, eds: Interaction of ultrasound and biological tissues workshop proceedings, DHEW Publication (FDA) 73-8008 BRH-DBE, U.S. Government Printing Office, Washington, DC, 1972.
2. O'Brien WD, Shore ML, Fred RK, Leach WM: On the assessment of risk to ultrasound. 1972 Ultrasonics Symposium Proceedings, edited by J. deKlerk, 486-490, IEEE Cat. No. 72 CHO 708-8 SU, New York, 1972.
3. Ulrick WD: Ultrasound dosage for nontherapeutic use on human beings - extrapolation from a literature survey. IEEE Trans Biomed Engr., BME-21, 48-51, 1974.
4. Wells PNT: The possibility of harmful biological effects in ultrasonic diagnosis. Cardiovascular Applications of Ultrasound. ed. by RS Reneman, 1-17, Elsevier, NY, 1974.
5. Hazzard DG, Litz ML, eds: Symposium on biological effects and characterization of ultrasound sources proceedings, DHEW Publication (FDA) 78-8048, U.S. Government Printing Office, Washington, DC, 1977.
6. O'Brien WD: Safety of ultrasound. Clinical Handbook of Ultrasound. ed. M deVlieger et al., 99-108, Wiley, New York, 1978.
7. Repacholi MH, Benwell DA, eds: Ultrasound short course transactions. Radiation Protection Bureau, Health Protection Branch, National Health and Welfare, Canada, 1979.



8. Repacholi MH: Ultrasound: Characteristics and biological action. National Research Council of Canada, Publication Number NRCC 19244, Ottawa, 1981.
9. Dunn F, Frizzell LA: Bioeffects of ultrasound. Therapeutic heat and cold, ed JF Lehmann, 386-403, Williams and Wilkins, Baltimore, 1982.
10. Stratmeyer ME, Stewart HF: An overview of ultrasound: theory, measurement, medical applications, and biological effects. HHS Publication FDA 82-8190, U.S. Government Printing Office, Washington, DC, 1982.
11. Environmental Health Criteria for Ultrasound. World Health Organization. This document should be available during 1983 through the WHO Geneva Office.
12. National Council on Radiation Protection and Measurement Document NCRP No. 66 should be available by the end of 1983 from NCRP, Washington, DC.
13. National Institute of Child Health and Human Development, NIH, Task Force on Use of Diagnostic Ultrasound Imaging in Pregnancy. The development of this document is through the NIH Consensus Development Conference process and is scheduled for completion in early, 1984.
14. Cady WG: Piezoelectricity. Vol. 1, New York, Dover Publication, Inc., 1946.
15. Urick RJ: Principles of underwater sound for engineers. New York, McGraw-Hill Book Co., 1967.

16. Van Went JM: Ultrasonic and ultrashort waves in medicine. New York, Elsevier Pub. Co., 1954.
17. Graber P: Biological actions of ultrasonic waves. Adv Biol Physics, eds., JH Lawrence, CA Tobias, vol. 3:191-246, Academic Press, New York, NY, 1953.
18. Wood RW, Loomis AL: The physical and biological effects of high-frequency sound-waves of great intensity. Phil Mag 4:417, 1927.
19. Harvey EN: Biological aspects of ultrasonic waves, A general survey. Biological Bulletin 59:306, 1930.
20. Fry WJ, Wulff VJ, Tucker D, et al: Physical factors involved in ultrasonically induced changes in living systems: I. Identification of non-temperature effects. J Acoust Soc Am 22:867, 1950.
21. Fry WJ, Tucker D, Fry JF, et al: Physical factors involved in ultrasonically induced changes in living systems: II. Amplitude duration relations and the effect of hydrostatic pressure for nerve tissue. J Acoust Soc Am 23:365, 1951.
22. Fry WJ: Intense ultrasound in investigation of the central nervous system. Adv Biol Med Physics 6:281, 1958.
23. Hueter TF, Ballantine HT Jr., Cotter WC: Production of lesions in the central nervous system with focused ultrasound. A study of dosage factors. J Acoust Soc Am 28:192, 1956.

24. Fry FJ, Kossoff G, Eggleton RC, Dunn F: Threshold ultrasonic dosages for structural changes in the mammalian brain. J Acoust Soc Am 48:1413-1417, 1970.
25. Pond JB: The role of heat in the production of ultrasonic focal lesions. J Acoust Soc Am 47:1607-1611, 1970.
26. Dunn F, Fry FJ: Ultrasonic threshold dosages for the mammalian central nervous system. IEEE Trans Bio-Med Eng. BME-18: 253-256, 1971.
27. Robinson TC, Lele PP: An analysis of lesion development in the brain and in plastics by high-intensity focused ultrasound at low-megahertz frequencies. J Acoust Soc Am 51:1333, 1972.
28. Dunn F: Physical mechanisms of the action of intense ultrasound on tissue. Am J Phys Med 37:148, 1958.
29. Taylor KJW, Pond J: The effects of ultrasound on varying frequencies on rat liver. J Path 100:287, 1969.
30. Eggleton RC, Kelly E, Fry FJ, et al: Morphology of ultrasonically irradiated skeletal muscle. Ultrasonic Energy. Kelly, E. (ed), Urbana, Illinois, University of Illinois Press, 117, 1965.
31. Ravitz MJ, Schnitzler RM: Morphological changes induced in the frog semitendinosus muscle fiber by localized ultrasound. Exptl Cell Res 60:78, 1970.
32. Fallon JT, Stephens WF: Effect of ultrasound on arteries.

Arch Path 94:380, 1972.

33. Frizzell LA, Linke CA, Carstensen EL, Fridd CW: Thresholds for focal ultrasonic lesions in rabbit kidney, liver and testicle. IEEE Trans Biomed Engr., BME-24, 393-396, 1977.
34. Colman DJ, Lizzi F, Burt W, et al: Ultrasonically induced cataract. Am J Ophthal 71:1284, 1971.
35. Sokollu A: Destructive effect of ultrasound on ocular tissue. JM Reid, M Sikov (eds), Interaction of Ultrasound and Biological Tissue, Washington, DC, US Gov't. Printing Office, DHEW Pub. (FDA) 73-8008, 1972, p. 129.
36. Lizzi FL, Parker AJ, Coleman DJ: Experimental cataract production by high frequency ultrasound. Ann Ophth 10:934-942, 1978.
37. Landau E: Are there ultrasonic danger for the unborn. Practical Radiology 1:27-31, 1973.
38. Roney PL, Albrecht RM: Hospital survey of obstetric ultrasound, presented at the Ad Hoc Review Panel on Ultrasound Bioeffect and Measurement meeting, Bureau of Radiological Health, FDA, April 9-10, 1976.
39. Roney PL, Albrecht RM: Hospital survey of obstetric ultrasound-United States, 1974. Symposium on Biological Effects and Characterization of Ultrasound Sources Proceedings, DG Hazzard and ML Litz (eds), HEW Publication (FDA) 78-8048, p. 29-30, US Government Printing Office, Washington, DC, 1977.

40. Ziskin MC: Survey of patient exposure to diagnostic. In JM Reid and MR Sikov (eds), Interaction of Ultrasound and Biological Tissues, DHEW Pub. (FDA) 73-8008, U.S. Government Printing Office, Washington, DC, 1972.
41. Wells PNT: What future of ultrasonics. Ultrasonics 11:16, 1973.
42. Smith SW: Diagnostic ultrasound: A review of clinical applications and the state of the art of commercial and experimental systems. HEW Publication (FDA) 76-8055, U.S. Government Printing Office, Washington, DC, 1976.
43. Smith SW: Diagnostic equipment and its use. Presented at the 8th Annual National Conference on Radiation Control, Springfield, IL, May 2-7, 1976.
44. Stratmeyer ME: Research directions in ultrasound bioeffects - A public health view. Symposium on Biological Effects and Characterizations of Ultrasound Sources Proceedings, DG Hazzard and ML Litz (eds), HEW Publication (FDA) 78-8048, p.240-248, US Government Printing Office, Washington, DC, 1977.
45. Frost HM, Stratmeyer ME: In-vivo effect of diagnostic ultrasound. The Lancet, p. 999, May 7, 1977.
46. Nyborg WL: Physical mechanisms for biological effects of ultrasound. HEW Publication (FDA) 78-8062, U.S. Government Printing Office, Washington, DC, 1978.

47. Nyborg WL: Physical mechanisms for biological effects of ultrasound. Ultrasound Short Course Transactions. MH Repacholi and DA Benwell (eds), 84-136, Radiation Protection Bureau, Health Protection Branch, National Health and Welfare, Canada, 1979
48. Dunn F: Biological effects of ultrasound. Ultrasound Short Course Transactions. MH Repacholi and DA Benwell (ed), 51-81, Radiation Protection Bureau, Health Protection Branch, National Health and Welfare, Canada, 1979.
49. Lele PP: Safety and potential hazards in the current applications of ultrasound in obstetrics and gynecology. Ultrasound Med Biol 5:307-320, 1979.
50. Anderson DW, Barrett JT: Ultrasound: A new immunosuppressant. Clinical Immunology Immunopathology 14:18-29, 1979.
51. Liebeskind D, Bases R, Elequin F, et al: Diagnostic ultrasound: Effects on the DNA and growth patterns of animal cells. Radiology 131:177-184, 1979.
52. Liebeskind D, Bases R, Mendex F, et al: Sister chromatic exchanges in human lymphocytes after exposure to diagnostic ultrasound. Science 205:1273-1275, 1979.
53. Siegel E, Goddard J, James AE, Siegel EP: Cellular attachment as a sensitive indicator of the effects of diagnostic ultrasound and exposure on cultured human cells. Radiology 133:175-179, 1979.

54. Miller DL, Nyborg WL, Whitcomb CC: Platelet aggregation induced by ultrasound under specialized conditions in vivo. *Science* 205:505-507, 1979.
55. Anderson DW, Barrett JT: Depression of phagocytosis by ultrasound. *Ultrasound Med Biol* 7:267-273, 1981.
56. Liebeskind D, Bases R, Koenigsberg M, Koss L, Raventos MS: Morphological changes in the surface characteristics of cultured cells after exposure to diagnostic ultrasound. *Radiology*, 138:419-423, 1981.
57. Haupt M, Martin AO, Simpson JL, Iqbal MA, Elias S, Dyer A, Sabbagha RE: Ultrasonic induction of sister chromatid exchanges in Human lymphocytes. *Hum Genet* 59:221-226, 1981.
58. Morris SM, Palmer CG, Fry FJ, Johnson LK: Effect of ultrasound on human leucocytes - sister chromatid exchange analysis. *Ultrasound Med Biol* 4:253-258, 1978.
59. Wegner RD, Obe G, Meyenburg M: Has diagnostic ultrasound mutagenic effects? *Hum. Gent.* 56:95-98, 1980.
60. Zheng HZ, Mitter NS, Chudley AE: In vivo exposure to diagnostic ultrasound and in vitro assay of sister chromatid exchanges in cultured amniotic fluid cells. *IRSC Med. Science: Biochemistry; Biomedical Technology; Cell and Membrane Biology; Clinical Biochemistry; Developmental Biology and Medicine; Environmental Biology and Medicine; Pathology; Reproduction; Obstetrics and Gynecology* 9:491, 1981.

61. Au WA, Obergoenner N, Goldenthal KL, Corry PM, Willingham V: Sister-chromatid exchanges in mouse embryos after exposure to ultrasound in utero. *Mutation Res* 103:315-320, 1982.
62. Barrass N, ter Haar G, Casey G: The effect of ultrasound and hyperthermia on sister chromatid exchange and division kinetics of BHK21 Cl3/A3 cells. *Brit. J. Cancer* 45 suppl v:187-191, 1982.
63. Lundberg M, Jerominski L, Livingston G, Kochenour N, Lee T, Findeman R: Failure to demonstrate an effect of in vivo diagnostic ultrasound on sister chromatid exchange frequency in amniotic fluid cells. *Am. J. Med. Genet.* 11:31-35, 1982.
64. Wegner RD, Meyenburg M: The effects of diagnostic ultrasonography on the frequency of sister chromatid exchanges in chinese hamster cells and human lymphocytes. *J. Ultrasound Med.* 1:355-358, 1982.
65. O'Brien WD Jr.: Ultrasonically induced fetal weight reduction in mice. *Ultrasound in Medicine.* eds D White and R Barnes p. 531-532, Plenum Press, New York, 1976.
66. O'Brien WD Jr., Christman CL, Yarrow S: Ultrasonic biological effect exposure system. 1974 Ultrasonic Symposium Proceedings, (ed) J deKlerk, p. 57-64, IEEE Catalog #74 CHO 896-ISU, New York, 1974.
67. O'Brien WD Jr: Dose-dependent effect of ultrasound on fetal weight in mice. *J Ultrasound Med* 2:1-8, 1983.



68. Dunn F, Lohnes JE, Fry FJ: Frequency dependence of threshold ultrasonic dosages for irreversible structural changes in mammalian brain. J Acoust Soc Am 58:512-514, 1975.
69. Johnston RL, Dunn F: Influence of subarachnoid structures on transmengeal ultrasonic propagation. J Acoust Soc Am 60:1225-1227, 1976.
70. Johnston RL, Dunn F: Ultrasonic absorbed dose, dose rate, and produced lesion volume. Ultrasonics 14:153, 1976.
71. Fry WJ, Dunn F: Ultrasonic irradiation of the central nervous system at high sound levels. J Acoust Soc Am 28:129-131, 1956.
72. Chan SK, Frizzell LA: Ultrasonic thresholds for structural changes in the mammalian liver. 1977 Ultrasonics Symposium Proceedings, J deKlerk, BR McAvoy (ed), IEEE Catalog no 77CH2364--ISU, New York, NY, 153-156, 1977.
73. Rossi HH, Kellere AM: Radiation carcinogenesis at low doses. Science 175:200-202, 1972.
74. Wang SY: Introductory concepts for photochemistry of nucleic acids. Photochemistry and Photobiology of Nucleic Acids, vol. 1 ed. SY Wang, p 1-21, Academic Press, New York, 1976.
75. Fry FJ, Erdmann WA, Johnson LK, Baird AI: Ultrasonic toxicity study. Ultrasound Med Biol 3:351-366, 1978.
76. Stolzenberg SC, Torbit CA, Edmonds PD, Taenzer JC: Effects of ultrasound on the mouse exposed at different stages of

gestation: Acute study. Radiation Environmental Biophysics  
17:245-270, 1980.

77. AIUM/NEMA Safety Standard for Diagnostic Ultrasound  
Equipment. AIUM/NEMA Standards Publication U11-1981.  
Available through the American Institute of Ultrasound in  
Medicine, Bethesda, MD or the National Electrical  
Manufacturers Association, Washington, DC, 1981.

TABLE 1

Selected Assessment of Diagnostic Ultrasound Usage

<u>Assessment Period</u>	<u>Assessment</u>
1963-1971	Annualized increase in useage <u>via</u> a mail survey about 10 percent (40).
early 1970's	In the United Kingdom, the number of diagnostic exams doubles every three years (26 percent annualized) (41).
1971	Twelve percent of hospitals use ultrasound (37).
1974	Thirty five percent of hospitals use ultrasound (38, 39).

TABLE 2

Summary of CF<sub>1</sub> mouse fetal weight study (65, 67). The mice were ultrasonically exposed (continuous wave of 1 MHz) in utero on day eight of gestation and the fetuses were examined on the eighteenth day of gestation.

<u>Isata (W/cm<sup>2</sup>)</u>	<u>t(sec)</u>	<u>I<sup>2</sup>t</u>	<u>Percentage weight change (re: sham)</u>
sham		0	0
0.5	300	75	- 5.3
2.0	20	80	- 6.1
3.0	10	90	- 6.1
0.7	300	147	- 8.8
3.0	20	180	- 7.9
5.5	10	303	-17.5

TABLE 3

Summary of LAF<sub>1</sub>/J mouse fetal weight study (<sup>75</sup>~~X~~). The mice were ultrasonically exposed (pulsed,  $I_{spta} = 1936 \text{ W/cm}^2$ , 1 MHz, pulsed repetition frequency = 1 kHz) in utero on the eighth day of gestation and the fetuses were weighed on the eighteenth day of gestation.

<u><math>I_{spta}</math> (W/cm<sup>2</sup>)</u>	<u>t(sec)</u>	<u><math>I^2t</math></u>	<u>Percentage weight change (re: sham)</u>
sham		0	0
25.6	20	13,100	- 5.4
35.3	20	24,900	0
45.0	20	40,500	0
50.8	20	51,600	-18.8

TABLE 4

Summary of CFW Swiss Webster mouse fetal weight study (<sup>76</sup>~~X1~~). The mice were ultrasonically exposed (continuous wave of 2 MHz) in utero on days 0, 7 or 12 of gestation and the fetuses were weighed on the seventeenth day of gestation.

<u>I</u> <sub>sata</sub> (W/cm <sup>2</sup> )	t(sec)	<u>I</u> <sup>z</sup> t	Percentage weight change (re: sham)		
			<u>day 0</u>	<u>day 7</u>	<u>day 12</u>
	sham	0	0	0	0
1	80	80	- 2.7	+ 5.1	- 3.9
1	100	100	+ 1.6	+ 3.9	- 5.6
1	120	120	- 0.4	- 0.4	- 5.3
1	140	140	+ 2.9	- 8.7	-13.3
1	160	160	- 1.8	-11.2	-22.2
1	180	180	-19.1	-11.0	-11.3
1	200	200	-25.1	-11.5	-18.4

# PERCENTAGE WEIGHT CHANGE (re : SHAM)

